



## POSTERS

Posters will be mounted in the Magpie, Wasatch & Superior Lobby and will be available for viewing from 12:00 PM, Monday, October 2, 2017. The posters will remain on display for the duration of the meeting and can be viewed until the specified times:

<b>Monday, October 2, 2017</b>	<b>8:30 PM</b>
<b>Tuesday, October 3, 2017</b>	<b>6:00 PM</b>
<b>Wednesday, October 4, 2017</b>	<b>1:10 PM</b>

During the poster sessions, Monday, October 2, 12:05 – 1:30PM (even numbers) & Monday, October 2, 6:30 – 8:30PM (odd numbers), the authors of the posters will be present and available for more in-depth discussion.

### List of Posters

#### **P-1 Comparative Gene Expression Analysis for Identification of Appropriate Markers of Pluripotency and Germline Competency**

Raymond R. Asuncion, Jennifer R Goldsmith, Anna Pham, Vida Asghari, Deborah Siler, Melanie Domeyer, J. Colin Cox, Xin Y Rairdan, Rhonda Wiler

Genentech, Inc., South San Francisco, CA, United States

#### **P-2 CRISPR/Cas9 mutant mouse model creation through in vitro fertilization and zygote electroporation**

Shinya Ayabe<sup>1</sup>, Tomohiro Tamari<sup>2</sup>, Kenichi Nakashima<sup>1</sup>, Eikichi Yanagisawa<sup>2</sup>, Mizuho Iwama<sup>1</sup>, Koji Nakade<sup>1</sup>, Takehide Murata<sup>1</sup>, Atsushi Yoshiki<sup>1</sup>, Yuichi Obata<sup>1</sup>

<sup>1</sup>RIKEN BioResource Center, Tsukuba, Ibaraki, Japan; <sup>2</sup>Charles River Laboratories Japan, Inc., Ishioka, Ibaraki, Japan

#### **P-3 Rat Resource and Research Center**

Elizabeth C Bryda, Hongsheng Men, Yuksel Agca, Aaron C Ericsson, James M Amos-Landgraf, Craig L Franklin

College of Veterinary Medicine, University of Missouri, Columbia, Missouri, United States

#### **P-4 Generation of interferon $\alpha/\beta$ receptor knockout sheep using CRISPR/Cas9 and SCNT techniques**

Zhiqiang Fan, Misha Regouski, Min Yang, Kirsten Gash, Qinggang Meng, Christopher J Davies, Young-Min Lee, Irina A Polejaeva

Utah State University, Logan, Utah, United States

### **P-5 RHEB1 insufficiency and stress-induced seizures in aged male mice**

Qi Tian<sup>1</sup>, P Gromov<sup>2</sup>, Y Wang<sup>1</sup>, J Clement<sup>3</sup>, M Riemann<sup>4</sup>, F Weih<sup>4</sup>, X Xin Sun<sup>1</sup>, M Shui Dai<sup>1</sup>, Lev Fedorov<sup>1</sup>

<sup>1</sup>Oregon Health & Science University, Portland, Oregon, United States; <sup>2</sup>Danish Cancer Society Research Center, Copenhagen, Denmark; <sup>3</sup>University Hospital, Jena, Germany; <sup>4</sup>Fritz Lipmann Institute, Jena, Germany

### **P-6 Generation of ES cell-derived mouse kidneys in anephric rats by blastocyst complementation**

Teppi Goto<sup>1</sup>, Hideki Masaki<sup>2</sup>, Hiromasa Hara<sup>1</sup>, Makoto Sanbo<sup>1</sup>, Hideyuki Sato<sup>2</sup>, Tomoyuki Yamaguchi<sup>2</sup>, Shinichi Hochi<sup>3</sup>, Hiromitsu Nakauchi<sup>4</sup>, Masumi Hirabayashi<sup>1</sup>

<sup>1</sup>National Institute for Physiological Sciences, Okazaki, Aichi, Japan; <sup>2</sup>The University of Tokyo, Tokyo, Japan; <sup>3</sup>Shinshu University, Ueda, Nagano, Japan; <sup>4</sup>Stanford University, Stanford, California, United States

### **P-7 Generation of knockout, knock-in, and humanized mouse models using the CRISPR/Cas9 technology: lessons learned and open questions**

Steffen Guettler<sup>1</sup>, Alexander Klimke<sup>1</sup>, Petric Kuballa<sup>1</sup>, Gesa Glöcke<sup>1</sup>, Jeanette Keßler<sup>1</sup>, Heidirun Kern<sup>1</sup>, Kenneth Albrecht<sup>2</sup>, Adriano Flora<sup>1</sup>, Eleanor Kolossovski<sup>2</sup>, Jochen Welcker<sup>1</sup>

<sup>1</sup>Taconic Biosciences GmbH, Cologne, Germany; <sup>2</sup>Taconic Biosciences Inc., Hudson, NY, United States

### **P-8 Complexity of the animals generated by Crispr**

Shuqin Zhang, Xiaohao Yao, Xianling Zhao, Shumei Zhao, Xin Su, Caiying Guo  
HHMI, Janelia Research Campus, Ashburn, VA, United States

### **P-9 Transfection of Mouse Pluripotent Stem Cells for Gene-Editing with CRISPR/CAS9 in a Feeder-Free Culture System**

James Kehler, Soojung Shin, Mark Kennedy, Joanna Asprer, David Kuninger  
Thermo Fisher Scientific, Frederick, MD, United States

### **P-10 Genome engineering using the Cre/loxP system**

Jason Klotz, Michael Grzybowski, Oleg Palygin, Aron M Geurts, Melinda Dwinell  
Medical College of Wisconsin, Milwaukee, WI, United States

### **P-11 A mouse model of MeCP2 overexpression**

Martha V Koerner, Jim Selfridge, Adrian P Bird  
University of Edinburgh, Edinburgh, United Kingdom

### **P-12 Towards the role of the epigenetic factor PRDM9 in meiosis of Rattus norvegicus**

Ondrej Mihola<sup>1</sup>, Vladimír Landa<sup>2</sup>, Petr Flachs<sup>1</sup>, Michal Pravenec<sup>2</sup>, Zdenek Trachtulec<sup>1</sup>

<sup>1</sup>Institute of Molecular Genetics AS CR, Prague, Czech Republic; <sup>2</sup>Institute of Physiology of the ASCR, Prague, Czech Republic



### **P-13 Novel Rodent Models of Renal Cystic Disease**

Hongsheng Men, Daniel J Davis, Kari L Chesney, Anagha S Bock, Miriam A Hankins, Elizabeth C Bryda

University of Missouri, Columbia, Missouri, United States

### **P-14 Using CRISPR to investigate the role of a placental-specific long non-coding RNA in mouse**

Laramie Pence, Jessica Martin, Bhanu Telugu

University of Maryland, College Park, MD, United States

### **P-15 Multiple Transgenic Rabbit Models for Human Diseases**

Xuwen Peng

Penn State University Hershey College of Medicine, Hershey, Pennsylvania, United States

### **P-16 RNAi and CRISPR/Cas9 based In Vivo Models for Drug Discovery**

Chia-lin Wang<sup>1</sup>, Yu-ting Yang<sup>1</sup>, Steven Sansing<sup>2</sup>, Christina Leslie<sup>3</sup>, Lukas Dow<sup>4</sup>, Johannes Zuber<sup>5</sup>, Scott W Lowe<sup>3</sup>, Prem K Premririt<sup>1</sup>

<sup>1</sup>Mirimus Inc., Brooklyn, NY, United States; <sup>2</sup>Charles River Laboratories, Wilmington, MA, United States;

<sup>3</sup>Memorial Sloan Kettering Cancer Center, New York, NY, United States; <sup>4</sup>Weill Cornell Medical College,

New York, NY, United States; <sup>5</sup>Research Institute of Molecular Pathology, Vienna, Austria

### **P-17 SMPD1 knockout golden Syrian hamsters develop spontaneous hyperlipidemia with elevated apolipoprotein B cholesterol**

Nick Robl, Rong Li, Zhongde Wang

Utah State University/Auratus Bio LLC., Logan, UT, United States

### **P-18 The phenotype of Taz null mutation in mouse closely resembles Barth Syndrome**

Douglas Strathdee<sup>1</sup>, Charlotte McCarroll<sup>1</sup>, Fiona Warrander<sup>2</sup>, Farah Hughes<sup>2</sup>, Frederic Vaz<sup>3</sup>, David Stevenson<sup>2</sup>, Laurence Cadalbert<sup>2</sup>, Sheila Bryson<sup>2</sup>, Nicola Laprano<sup>2</sup>, Christopher M Loughrey<sup>1</sup>, Eve Anderson<sup>2</sup>

<sup>1</sup>University of Glasgow, Glasgow, Scotland, United Kingdom; <sup>2</sup>CRUK Beatson Institute, Glasgow, Scotland, United Kingdom; <sup>3</sup>Academic Medical Center, Amsterdam, Netherlands

### **P-19 Efficient use of vitrified or refrigerated pronuclear zygotes for gene-modification in rats with CRISPR/Cas9 system**

Akiko Takizawa<sup>1</sup>, Tomoo Eto<sup>2</sup>, Lynn Lazcares<sup>1</sup>, Rebecca Schilling<sup>1</sup>, Michael Grzybowski<sup>1</sup>, Aron Geurts<sup>1</sup>, Masumi Hirabayashi<sup>3</sup>, Melinda R Dwinell<sup>1</sup>

<sup>1</sup>Medical College of Wisconsin, Milwaukee, WI, United States; <sup>2</sup>Central Institute for Experimental Animals, Kawasaki, Kanagawa, Japan; <sup>3</sup>National Institute for Physiological Sciences, Okazaki, Aichi, Japan

### **P-20 Oxidative stress-induced tumorigenesis: Lesson from the experiments with DNA repair-deficient mice**

Teruhisa Tsuzuki<sup>1</sup>, Mizuki Ohno<sup>1</sup>, Noriko Takano<sup>1</sup>, Ken-ichi Taguchi<sup>2</sup>, Yusaku Nakabeppu<sup>3</sup>, Yoshimichi Nakatsu<sup>1</sup>

<sup>1</sup>Kyushu University, Fukuoka, Japan; <sup>2</sup>National Hospital Organization, Kyushu Cancer Center, Fukuoka, Japan; <sup>3</sup>Medical Institute of Bioregulation, Kyushu University, Fukuoka, Japan

### **P-21 The naturally short-lived turquoise killifish sheds light on the basis of vertebrate lifespan**

Dario Riccardo Valenzano

*Max Planck Institute for Biology of Ageing, Cologne, Germany*

### **P-22 Off-target Analysis of CFTR-Knockout Sheep Produced by CRISPR/Cas9 and Somatic Cell Nuclear Transfer**

Iuri Viotti Périssé\*, Zhiqiang Fan\*, Zhongde Wang, Kenneth L White, Irina A Polejaeva  
Utah State University, Logan, UT, United States \* these authors contributed equally to this project.

### **P-23 Modifying the Opossum genome using CRISPR**

Sophie Wood, Fanny Decarpentrie

*Francis Crick Institute, London, UK*

### **P-24 Cav 1.2 sequence and mutually exclusive splicing study to develop pig models of cardiac and neuropsychiatric disorders**

Elisa Zanfrini<sup>1</sup>, Andrea Perota<sup>1</sup>, Irina Lagutina<sup>1</sup>, Constanza Lopez<sup>1</sup>, Roberto Duchi<sup>1</sup>, Silvia Colleoni<sup>1</sup>, Silvia Fasciano<sup>2</sup>, Andrea Mazzanti<sup>4</sup>, Giovanna Lazzari<sup>1,3</sup>, Silvia G. Priori<sup>2,4</sup>, Cesare Galli<sup>1,3</sup>

<sup>1</sup>Avantea, Laboratory of Reproductive Technologies, Cremona, Italy; <sup>2</sup>Department of Cardiology, University of Pavia, Pavia, Italy; <sup>3</sup>Fondazione Avantea, Cremona, Italy; <sup>4</sup>Department of Molecular Cardiology, IRCCS Fondazione Salvatore Maugeri, Pavia, Italy

### **P-25 Joint Intervention of E-cadherin and Bmi-1 Genes Mediated**

Tingting Luo#, Aifen Yan#, Hong Jiang, Lian Liu, Cuilan Feng, Fang Liu, Dongsheng Tang  
Foshan University, Foshan, Guangdong, China # Co-First authors

### **P-26 CRISPR/Cas9 mediated genome editing by electroporation of mouse oocytes**

Fabien Delerue, Lars M Ittner

*University of New South Wales, SYDNEY, New South Wales, Australia*

### **P-27 A method to convert mRNA into a gRNA library for CRISPR/Cas9 editing of any organism**

Hiroshi Arakawa

*Institute of Molecular Oncology Foundation, Milan, Italy*



TT2017

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### **P-28 Maximizing certainty of ES cell germline transmission rates - A cascade of efficiencies using genetically sterile "goGermline" embryos**

Roger Askew

Ozgene

### **P-29 TARGETED GENE REPLACEMENT OF 42kb USING ASYMMETRIC DONORS IN mES CELLS**

Oliver Baker<sup>1</sup>, Sarah Tsurkan<sup>2</sup>, Jun Fu<sup>2</sup>, Barbara Klink<sup>3</sup>, Andreas Rump<sup>3</sup>, Mandy Obst<sup>2</sup>, Andrea Kranz<sup>2</sup>, Evelin Schroeck<sup>3</sup>, Konstantinos Anastassiadis<sup>4</sup>, Francis A Stewart<sup>2</sup>

<sup>1</sup>King's College London, London, UK; <sup>2</sup>Genomics, Biotechnology Center, Technische Universität Dresden, Germany; <sup>3</sup>Institute for Clinical Genetics, Faculty of Medicine, Carl Gustav Carus, Technische Universität Dresden, Germany; <sup>4</sup>Stem Cell Engineering, Biotechnology Center, Technische Universität Dresden, Germany

### **P-30 Site-specific transgene insertion mediated by CRISPR/Cas9 and homology-directed DNA repair pathway at the H11 and ROSA26 loci in bovine fetal fibroblast cells**

Cicera R Lazzarotto<sup>1</sup>, Carlos Pinzon<sup>2</sup>, Hanah Georges<sup>2</sup>, Marcelo Bertolini<sup>3</sup>, Charles R Long<sup>2</sup>, Luciana R Bertolini<sup>1</sup>

<sup>1</sup>Pontifical Catholic University of Rio Grande do Sul, Brazil; <sup>2</sup>Texas A&M University, College Station, Texas, United States; <sup>3</sup>Federal University of Rio Grande do Sul, Brazil

### **P-31 Genomic rearrangements generated by CRISPR-Cas9 and their characterization pitfalls**

Katharina M Boroviak, Beiyuan Fu, Graham Duddy, Brendan Doe, Edward Ryder, Fengtang Yang, Allan Bradley

Wellcome Genome Campus, Cambridge, Cambridgeshire, United Kingdom

### **P-32 " Replications, ridicule and a recluse " NgAgo and beyond**

Nay Chi Khin, Jenna Lowe, Lora Starrs, Gaetan Burgio

The Australian National University, Canberra, ACT, Australia

### **P-33 CRISPR/Cas9 for in vivo gene editing: the hidden face of paradise**

Marie-Christine Birling<sup>1</sup>, Laurence Schaeffer<sup>1</sup>, Philippe Andre<sup>1</sup>, Loic Lindner<sup>1</sup>, Jimmy Mancip<sup>2</sup>, Abdelkader Ayadi<sup>1</sup>, Tania Sorg<sup>1</sup>, Yann Herault<sup>1</sup>, Jean Cozzi<sup>2</sup> & Guillaume Pavlovic<sup>1</sup>

<sup>1</sup>PHENOMIN, Institut Clinique de la Souris (ICS), CNRS, INSERM, University of Strasbourg, 1 rue Laurent Fries, F-67404 Illkirch-Graffenstaden, France; <sup>2</sup>Charles River, European Embryology Platform, BP 109 - Domaine Des Oncins, 69210 Saint-Germain-Nuelles, France

### **P-34 Induced- pluripotent stem cells from transgenic mice NF-κB-RE-Luc: First steps in the generation of new reporter cell models for the study of inflammation**

Romina Pagotto<sup>1</sup>, Micaela Sureda<sup>1,2</sup>, Belen Harreguy<sup>1</sup>, Geraldine Schlapp<sup>2</sup>, Martina Crispo<sup>2</sup>, Mariela Bollati-Fogolin<sup>1</sup>

<sup>1</sup>Cell Biology Unit, Institut Pasteur de Montevideo, Uruguay; <sup>2</sup>Transgenic and Experimental Animal Unit, Institut Pasteur de Montevideo, Uruguay

### **P-35 Generation and characterization of a stable transgenic ZsGreen reporter rat line**

Daniel J Davis, Hongsheng Men, Anagha S Bock, Mary L Shaw, Kari L Chesney, Miriam A Hankins, Elizabeth C Bryda  
University of Missouri, Columbia, Missouri, United States

### **P-36 Targeted insertion of polyA tracks with CRISPR-Cas9 allows titratable control of gene expression**

Rachel B. Delston<sup>1</sup>, Edward J Weinstein<sup>1</sup>, Xiaoxia Cui<sup>2</sup>, Yi-Hsien Chen<sup>2</sup>, Lucy Liaw<sup>3</sup>, Laura A Arthur<sup>2</sup>, Sergej Djuranovic<sup>2</sup>

<sup>1</sup>Canopy Biosciences, Saint Louis, Missouri, United States; <sup>2</sup>Washington University in Saint Louis, Saint Louis, Missouri, United States; <sup>3</sup>Maine Medical Center Research Institute, Scarborough, Maine, United States

### **P-37 C57BL/6NCrl mouse models generated via CRISPR/Cas9-mediated gene-editing**

Astrid Jensen<sup>1</sup>, Christopher Dowdy<sup>2</sup>, Don Liu<sup>2</sup>, Lieke Geerts<sup>1</sup>, Jeroen DeGroot<sup>1</sup>, Uma Saha<sup>2</sup>, Greg Martin<sup>2</sup>, Anne-Marie Zuurmond<sup>1</sup>, Stephen Festin<sup>2</sup>

<sup>1</sup>Charles River Leiden, Leiden, Netherlands; <sup>2</sup>Charles River Wilmington, Wilmington, MA, United States

### **P-38 Replacing a transgene with another one in transgenic mice by CRISPR/Cas9 technology**

Ferenc Erdélyi, Z Máté, É Kriván, K Kovács, N Hájos, G Szabó  
Institute of Experimental Medicine, Budapest, Hungary

### **P-39 Inducible regulation of ROR $\gamma$ t in mice with a modified reverse tetracycline-controlled transcriptional silencer 'rtTS3'**

Yasuyuki Fusamae, Tomohiko Okuda  
SHIONOGI & CO., LTD., Suzuki, Japan

### **P-40 Can the RNA-guided AsCpf1 nuclease be useful for generation of knock-in mice?**

Dawn E. Watkins-Chow<sup>1</sup>, Lisa J. Garrett<sup>2</sup>

Genetic Disease Research Branch and <sup>2</sup>Embryonic Stem Cell and Transgenic Mouse Core, National Human Genome Research Institute, NIH Bethesda, Maryland, United States

### **P-41 The Embryo Cradle: A Novel and Versatile Tool for Manipulating Mouse Embryos**

Teresa M Gunn<sup>1</sup>, Paul Taylor<sup>2</sup>, Jesse Gavin<sup>2</sup>

<sup>1</sup>McLaughlin Research Institute, Great Falls, MT, United States; <sup>2</sup>GeneSearch, Inc., Bozeman, MT, United States



TT2017

## 14TH TRANSGENIC TECHNOLOGY MEETING

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### **P-42 Implementing novel technologies in a transgenic facility: creating conditional alleles with CRISPR/Cas9 in mouse zygotes and embryonic stem cells**

Colin Pritchard<sup>1</sup>, Lona Kroese<sup>1</sup>, Rahmen Bin Ali<sup>1</sup>, Tanya M. Braumuller<sup>1</sup>, Linda Henneman<sup>1</sup>, Jessica Del Bravo<sup>1</sup>, Fina van der Ahe<sup>1</sup>, Huub van Vugt<sup>2</sup>, Margriet Snoek<sup>2</sup>, Ivo J. Huijbers<sup>1</sup>

<sup>1</sup>MCCA Transgenic Facility, The Netherlands Cancer Institute, Amsterdam, The Netherlands;

<sup>2</sup>Division of Molecular Genetics, The Netherlands Cancer Institute, Amsterdam, The Netherlands

### **P-43 Efficient homology-directed repair using single-stranded DNA templates**

Ashley M Jacobi, Christopher Vakulskas, Michael A Collingwood, Garrett R Rettig, Mark A Behlke

Integrated DNA Technologies, Inc., Coralville, IA, United States

### **P-44 Analysis of mouse genome changes after CRISPR/Cas9 editing in vivo**

Yingming Wang, Qi Tian, Lev Fedorov

Oregon Health & Science University, Portland, Oregon, USA

### **P-45 Application of Oocyte-Cytoplasmic Injection for CRISPR-Cas9 Gene-Editing in Mouse**

Sang Yong Kim<sup>1</sup>, Lu Yang<sup>1</sup>, So Young Kim<sup>2</sup>, Keith Gembala<sup>3</sup>, Sergei Korolov<sup>4</sup>, David Levy<sup>5</sup>

<sup>1</sup>New York University School of Medicine, New York, NY, United States;

<sup>2</sup>NYU Langone Medical Center, Division of Advanced Research Technologies (DART) and Rodent Genetic Engineering (RGE) Laboratory, New York, NY, United States; <sup>3</sup>Morrell Instrument Company, New York, NY, United States; <sup>4</sup>New York University School of Medicine, Department of Pathology, New York, NY, United States; <sup>5</sup>New York University School of Medicine, Department of Pathology and Molecular Pathology, New York, NY, United States

### **P-46 Comparison of Standardized Nomenclature for CRISPR, ZFN, TALEN Mutations Across Species**

Michelle N Knowlton, Cynthia L Smith

The Jackson Laboratory, Bar Harbor, ME, United States

### **P-47 STREAMLINING THE GENETICS OF THE LABORATORY RAT THROUGH GENOME EDITING AND ASSISTED REPRODUCTIVE TECHNOLOGIES**

Kathy J Krentz

University of Wisconsin, Madison, WI, United States

### **P-48 Knocking Out Recombineering: An Innovative and Rapid Approach for the Targeted Generation of Complex Allelic Modifications**

Gary Kucera, Cheryl Bock, Michael Flores, Meilang Flowers, Julie Kent, Ozge Kud-dar, Scott Soderling

Duke University, Durham, North Carolina, United States

**P-49 Highly efficient screening method for founders of CRISPR/Cas9-generated conditional knockout mice using full-length PCR**

I-Shing Yu<sup>2</sup>, Tzu-Shan Huang<sup>1</sup>, Tzu-Wen Wang<sup>1</sup>, Chien-Hui Wu<sup>1</sup>, Hsiao-Ying Yu<sup>1</sup>, Shen-Wen Wang<sup>1</sup>, Cheng-Ju Wang<sup>1</sup>, Bo-Yueh Wu<sup>1</sup>, Chun-Yu Chen<sup>1</sup>, Shu-Wha Lin<sup>1,3,4</sup>  
<sup>1</sup>Department of Clinical Laboratory Sciences and Medical Biotechnology, College of Medicine, National Taiwan University, Taipei, Taiwan; <sup>2</sup>Laboratory Animal Center, College of Medicine, National Taiwan University, Taipei, Taiwan; <sup>3</sup>Center of Genomic Medicine, National Taiwan University, Taipei, Taiwan; <sup>4</sup>Department of Laboratory Medicine, National Taiwan University Hospital, College of Medicine, National Taiwan University, Taipei, Taiwan

**P-50 Four-dimensional phenotypic analyses of mouse and human kidney development**

Nils O Lindstrom, Andrew P McMahon  
University of Southern California, Los Angeles, California, United States

**P-51 Applying CRISPR/Cas9 technology to the generation of genetically modified rodent models**

Evguenia Kouranova, Zhenyi Liu, and Kevin Forbes  
Horizon Discovery, St. Louis, MO, United States

**P-52 CRISPR/Cas9 genome editing, from plasmid DNA microinjection to dual guide/RNP electroporation: evolution of a small core facility for transgenic mice**

Marie Teixeira<sup>1</sup>, Valérie Risson<sup>1,3</sup>, Farida Henry<sup>1</sup>, Agnès Couzon<sup>1,2</sup>, Denise Aubert<sup>1,2</sup>, Frédéric Flamant<sup>2</sup>, Suzy Markossian<sup>1,2</sup>  
<sup>1</sup>SFR BioSciences, Plateau de Biologie Expérimentale de la Souris (AniRA-PBES), Ecole Normale Supérieure de Lyon, Université Lyon 1, CNRS UMS3444, INSERM US8, France; <sup>2</sup>Institut de Génomique Fonctionnelle de Lyon, Ecole Normale Supérieure de Lyon, Université Lyon 1, CNRS UMR5242, INRA USC1370, France; <sup>3</sup>Institut Neuro-MyoGène, Ecole Normale Supérieure de Lyon, Université Lyon 1, CNRS UMR5310, INSERM U1217, France

**P-53 Derivation and profiling of mouse epiblast stem cell lines (mEpiSCs)**

Javier Martin Gonzalez<sup>1</sup>, Xiaogang Guo<sup>2</sup>, Niels Alvaro Menezes<sup>2</sup>, Luca Mariani<sup>2</sup>, Elisabetta Ferretti<sup>2</sup>  
<sup>1</sup>Transgenic Core Facility, Department of Experimental Medicine, University of Copenhagen, Denmark; <sup>2</sup>The Danish Stem Cell Centre, Danstem, University of Copenhagen, Denmark

**P-54 CRISPR-mediated mutagenesis in cells and embryos**

Charleen Hunt, Anthony Gagliardi, Susannah Brydges, Gustavo Droguett, Jade Zhang, Jadine Vallelunga, Clarissa Herman, Michael Kelley, Amanda Karr, Lynn Lee, Katie Huling, Jacob Boysen, Yajun Tang, Tim Hanna, Jacqueline Buckley, Ella Dragileva, Lakeisha Esau, Sean Trzaska, Janelle Colon, Julita Posca, Jean Siao, Marine Prissette, William Poueymirou, David Frendewey, Wojtek Auerbach, Guochun Gong, Brian Zambrowicz  
Regeneron Pharmaceuticals Inc.





### **P-55 Generation of Hypoxanthine-guanine Phosphoribosyltransferase Gene Knockout Pigs by Somatic Cells Nuclear Transfer**

Haitao Wang, Xiaojuan Liu; Xiaoxiang Hu, Ning Li

State Key Laboratory for Agrobiotechnology, College of Biological Sciences, China Agricultural University, Beijing, China

### **P-56 Preliminary Characterization of p53 Knockout Golden Syrian Hamster Created by CRISPR/Cas9 System**

Jinxin Miao<sup>1,2</sup>, Rong Li<sup>1</sup>, Yaohe Wang<sup>2,3</sup>, Zhongde Wang<sup>1</sup>

<sup>1</sup>Department of Animal, Dairy, and Veterinary Sciences, Utah State University, Logan, Utah, USA; <sup>2</sup>National Centre for International Research in Cell and Gene Therapy, Sino-British Research Centre, School of Basic Medical Sciences, Academy of Medical Sciences, Zhengzhou University, Zhengzhou, P. R. China; <sup>3</sup>Centre for Molecular Oncology, Barts Cancer Institute, Queen Mary University of London, London, UK

### **P-57 Overall evaluation of gene modified mouse production by embryo-based genome editing**

Kanako Kato<sup>1</sup>, Saori Mizuno-Iijima<sup>1</sup>, Yoshihisa Ikeda<sup>2</sup>, Yoko Daitoku<sup>1</sup>, Yoshikazu Hasegawa<sup>1</sup>, Shinya Ayabe<sup>3</sup>, Yoko Tanimoto<sup>1</sup>, Ken-ichi Yagami<sup>1</sup>, Satoru Takahashi<sup>1</sup>, Fumihiro Sugiyama<sup>1</sup>

<sup>1</sup>University of Tsukuba, Tsukuba, Ibaraki, Japan; <sup>2</sup>Charles River Laboratories Japan, Ishioka, Ibaraki, Japan; <sup>3</sup>RIKEN BioResource Center, Tsukuba, Ibaraki, Japan

### **P-58 Generation of new CRISPR-edited mouse models for investigating non-syndromic types of albinism**

Santiago Josa, Almudena Fernández, Davide Seruggia, Andrea Montero, Yaiza López, Celia de Lara, Marcos Rubio, Iván Caballero, Diego Muñoz, Julia Fernández, Marta Cantero, Lluís Montoliu

CNB-CSIC and CIBERER-ISCIII, Madrid, Spain

### **P-59 Use of vitrified/warmed zygotes by spatula-MVD to generate CRISPR/Cas9 conditional knockout mouse**

Ana P Mulet, María N Meikle, Geraldine Schlapp, Martina Crispo

Institut Pasteur de Montevideo, Montevideo, Montevideo, Uruguay

### **P-60 improved GONAD (i-GONAD) (I) as ex vivo manipulation-free genome-editing system allowing efficient knock-out, large deletion, and knock-in**

Masato Ohtsuka<sup>1</sup>, Hiromi Miura<sup>1</sup>, Channabasavaiah B Gurumurthy<sup>2</sup>, Masahiro Sato<sup>3</sup>

<sup>1</sup>Tokai University, Isehara, Kanagawa, Japan; <sup>2</sup>University of Nebraska Medical Center, Omaha, Nebraska, United States; <sup>3</sup>Kagoshima University, Kagoshima, Kagoshima, Japan

### **P-61 A "gene editing journey" from gene targeting to CRISPR: impact of CRISPR/Cas in the generation of genetically modified mice**

Jaime Muñoz<sup>1</sup>, Pierfrancesco Vargiu<sup>1</sup>, Lucía Pérez de Ayala<sup>1</sup>, Martina Reiss<sup>2</sup>, Estefanía Ayala<sup>1</sup>, Harald Kranz<sup>2</sup>, Sagrario Ortega<sup>1</sup>

<sup>1</sup>Spanish National Cancer Research Centre (CNIO), Madrid, Spain; <sup>2</sup>Gene Bridges GmbH, Heidelberg, Germany

**P-62 Lmx1b-Dre, a CRISPR generated knock-in mouse for Dre mediated labelling of excitatory dorsal horn neurons**

Pelczar Pawel<sup>1</sup>, Wildner<sup>2</sup>, Oller<sup>1</sup>, Zeilhofer<sup>2</sup>

<sup>1</sup>University of Basel, Basel, Switzerland; <sup>2</sup>University of Zurich, ETH Zurich, Zurich, Switzerland

**P-63 V2G locus editing for production of GLP1Ser8-M3R transgenic pig islets**

Andrea Perota<sup>1</sup>, Irina Lagutina<sup>1</sup>, Roberto Duchi<sup>1</sup>, Nizar Mourad<sup>2</sup>, Elisa Zanfrini<sup>1</sup>, Constanza Lopez<sup>1</sup>, Silvia Colleoni<sup>1</sup>, Giovanna Lazzari<sup>1,3</sup>, Pierre Gianello<sup>2</sup> and Cesare Galli<sup>1,3</sup>

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**P-64 Screening for off-target CRISPR Mutations: It's never too late**

Anna Pham, Deborah Siler, Maria Martinez, Emily Hunley, J. Colin Cox

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**P-65 Genome editing in Rats: generation of a target point mutation by Pro-nuclear or Cytoplasmic Injection or by Electroporation of CRISPR/Cas9**

Sandra Turon<sup>1</sup>, Anna Arbo's<sup>1</sup>, Joan Bertolin<sup>2</sup>, Fa'tima Bosch<sup>2</sup>, Anna Pujol<sup>1</sup>

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**P-66 Improved GONAD (i-GONAD) (II): usefulness of rhodamine-dextran for monitoring the success of the GONAD and of gonadotrophin-based regulation of the timing for the GONAD**

Masahiro Sato<sup>1</sup>, Shingo Nakamura<sup>2</sup>, Channabasavaiah B Gurumurthy<sup>3</sup>, Satoshi Watanabe<sup>4</sup>, Masato Ohtsuka<sup>5</sup>

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**P-67 CRISPR/Cas9 Genome Editing Pipeline for Mice and Rats**

Thom L Saunders, Wanda E Filipiak, Galina B Gavrilina, Anna K La Forest, Corey E Ziebell, Michael G Zeidler, Elizabeth D Hughes

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**P-68 Introduction of Targeted Point Mutations in Mice using CRISPR-Cas9**

William Shawlot, Jinxiang Ren

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**P-69 Precision Medicine in Action: Using CRISPR/Cas9 to generate a human disease SNP rat model by electroporation of a single stranded DNA oligo**

Leslie A Stewart<sup>1</sup>, Brandon C Willis<sup>1</sup>, Diana L Young<sup>1</sup>, Nichole L Anchell<sup>1</sup>, Kayla M Jager<sup>1</sup>, Phuong T Dao<sup>1</sup>, Kristin N Grimsrud<sup>1</sup>, Joshua A Wood<sup>1</sup>, Kent C Lloyd<sup>1,2</sup>, Hermann Zbinden-Foncea<sup>4</sup>, Keith Baar<sup>3</sup>

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**P-70 Multi-gene Combined Intervention of Metastasis in Nasopharyngeal Carcinoma mediated by Gene Targeting with High Efficiency**

Dongsheng Tang<sup>1#</sup>, Wenzhu Huang<sup>2#</sup>, Tingting Luo, Hong Jiang, Cuilan Feng, Aifen Yan  
1Medical School, Foshan University, Foshan, Guangdong, China <sup>2</sup>Foshan Fifth People's Hospital, Foshan University, Foshan, Guangdong, China # Co-First authors

**P-71 Generation of TP53-modified pigs by GEEP method: CRISPR/Cas9-mediated gene modification introduced into porcine zygotes by electroporation**

Fuminori Tanihara<sup>1</sup>, Do Thi Kim Lanh<sup>1</sup>, Thanh-Van Nguyen<sup>1</sup>, Toshiki Kunihara<sup>1</sup>, Katsutoshi Nishio<sup>1</sup>, Tatsuya Takemoto<sup>2</sup>, Takeshige Otoi<sup>1</sup>

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**P-72 Extending the range of alleles obtained using CRISPR/Cas9 technology in mouse embryos: Generation and validation of mutations**

Gemma Codner, Francesca Pike, Joffrey Mianné, Adam Caulder, Ruairidh King, Rachel Fell, Alasdair Allan, Jorik Loeffler, Skevoulla Christou, Christopher McCabe, MLC Gene Delivery team, Sara Wells, Lydia Teboul  
MRC Harwell Institute, Didcot, Oxon, UK

**P-73 IN-DEPTH ANALYSIS OF CRISPR OFF-TARGETS IN GENETICALLY ENGINEERED RODENTS**

Keith R Anderson<sup>1</sup>, Maximillian Haussler<sup>2</sup>, Colin Watanabe<sup>1</sup>, Steffen Durinck<sup>1</sup>, Charles Yu<sup>1</sup>, Sobha R Thamminana<sup>1</sup>, Lucinda Tam<sup>1</sup>, Tuija Alcantar<sup>1</sup>, Jinjie Li<sup>1</sup>, Natasha O'Neil<sup>1</sup>, Xin Rairdan<sup>1</sup>, Merone Roose-Girma<sup>1</sup>

<sup>1</sup>Genentech, inc, South San Francisco, CA, United States; <sup>2</sup>UCSC, Santa Cruz, CA, United States

**P-74 Targeting human SerpinA1 by Cas9 therapeutic gene editing reverts liver damage phenotypes in a humanized  $\alpha$ 1-antitrypsin disease mouse model**

Mikael Bjursell<sup>1</sup>, Michelle J Porritt<sup>1</sup>, Elke Ericson<sup>1</sup>, Amir Taheri-Ghahfarokhi<sup>1</sup>, Maryam Clausen<sup>1</sup>, Lisa Magnusson<sup>1</sup>, Therese Admyre<sup>1</sup>, Roberto Nitsch<sup>1</sup>, Lorenz Mayr<sup>1</sup>, Leif Aasehaug<sup>2</sup>, Frank Seeliger<sup>2</sup>, Marcello Maresca<sup>1</sup>, Mohammad Bohlooly-Y<sup>1</sup>, John Wiseman<sup>1</sup>

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**P-75 Efficient generation of genetically edited rodent models using CRISPR-Cas9 by high throughput electroporation of zygotes**

Brandon C. Willis<sup>1</sup>, K. C. Kent Lloyd<sup>1,2</sup>, Joshua A. Wood<sup>1</sup>

<sup>1</sup>Mouse Biology Program; <sup>2</sup>Dept. of Surgery, School of Medicine, University of California, Davis

**P-76 Single strand DNA-mediated knock-in for large genomic regions in rodents**

Kazuto Yoshimi, Yayoi Kunihiro, Tomoji Mashimo

Genome Editing Research and Development Center, Graduate School of Medicine, Osaka University

**P-77 Efficient generation of gene knockout rats using CRISPR/Cas9**

Hee-Sook Bae<sup>1</sup>, Yun-Kyeong Jin<sup>2</sup>, Ok-Jae Koo<sup>1</sup>, JaeYoung Lee<sup>1</sup>, Goo Jang<sup>2</sup>

<sup>1</sup>Toolgen Inc. Seoul, Republic of Korea; <sup>2</sup>Department of veterinary clinical science, College of Veterinary Medicine, Seoul National University, Republic of Korea

**P-78 Whole genome analysis of a germline transmitted calf by next generation sequencing**

Soo-Young Yum<sup>1</sup>, Sinki Park<sup>2</sup>, SongJeon Lee<sup>3</sup>, Hee-Soo Kim<sup>3</sup>, Hyeong-Jong Kim<sup>3</sup>, Yun-Kyeong Jin<sup>1</sup>, Ji-Hyun Lee<sup>1</sup>, Goo Jang<sup>1</sup>

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**P-79 A new, easy and reliable cryopreservation protocol for zebrafish sperm**

Sandra Martins, Ana Catarina Certal

Chamalimaud Foundation, Lisbon, Portugal

**P-80 Enhanced CRISPR-mediated gene knockin by delaying zygote cell cycle progression with low temperature**

Tzu-Yu Chou, Mei-Ling Chang, Pi-Fang Tsai and Ching-Yen Tsai

Transgenic Core Facility, Inst. of Molecular Biology, Academia Sinica, Taiwan

**P-81 Efficient CRISPR reagent generation and genome engineering**

Xiaoxia Cui, Monica Sentmanat, Yi-Hsien Chen, Mike White, Colin Florian

Washington University in St. Louis, St. Louis, MO, United States

**P-82 Sperm Cryopreservation of Genetically Engineered Mouse Models: Why Quality Control by In Vitro Fertilization is Important?**

Amarnath Dasari, Andrea Foley, Susan Vanvliet, Sarah Erickson, Rosemary Leonard

Taconic Bio Sciences, Inc., Hudson, NY, United States

**P-83 Genetics meets 3 R's: Genetic background of transgenic mouse strains**

Peter Dobrowolski, Melina Fischer, Olaf Gelsen

GVG Genetic Monitoring GmbH, Leipzig, Germany

**P-84 A comparison of pseudopregnant F1c**

Timothy Corbin, Michael H Durnin, Rodney McCay, Jessica McCann, Heather Marshall

Stowers Institute for Medical Research, Kansas City, Missouri, United States

**P-85 Optimized co-transfection of murine ES cells**

Victor Badillo Lisakowski, Annette C. Fuchtbauer, Ernst-Martin Fuchtbauer

Department of Molecular Biology and Genetics, Aarhus University, Aarhus, Denmark



**P-86 Genome editing at TIGM: Production of various mutant alleles via CRISPR/Cas9**

Andrei Golovko, Huiping Guo, Johnathan Ballard, Amy Gonzales, John Adams, Benhamin Morpurgo

Texas A&M Institute for Genomic Medicine, College Station, TEXAS, United States

**P-87 Electroporation of Rat Embryonic Fibroblasts to Assess Function of CRISPR-Cas9 Plasmids**

Elizabeth D Hughes<sup>1</sup>, Wanda Filipiak<sup>2</sup>, Michael Zeidler<sup>1</sup>, Thomas L Saunders<sup>1</sup>

<sup>1</sup>BRCF-Transgenic Animal Model Core, University of Michigan, Ann Arbor, MI; <sup>2</sup>University of Michigan, Ann Arbor, MI, United States

**P-88 Validating custom and imported rodent lines during colony creation with PCR, qPCR and ddPCR**

Emily Hunley, Deborah Siler, Carol Cain-Hom, Anna Pham, Maria Martinez, Sanjeet Aujla, Rhonda Wiler, J Colin Cox

Transgenic Technologies, Genentech, South San Francisco, California

**P-89 Precision genome editing in the chicken using the CRISPR/Cas9 system**

Alewo Idoko Akoh, Helen Sang, Mike McGrew

University of Edinburgh, Easter Bush, Midlothian, Scotland, United Kingdom

**P-90 Generation of Novel Neurofibromatosis Type I Rodent Models**

Laura J Lambert, PhD, Anil K Challa, PhD, Ashley NTurner, MS, Daniel Kennedy, Bruce R Korf, MD, PhD, Robert A Kesterson, PhD

University of Alabama at Birmingham, Birmingham, AL, United States

**P-91 Harnessing the one-step generation of genetically engineered mice with CRISPR/Cas9 in zygotes**

Hiroshi Kiyonari<sup>1,2</sup>, Takaya Abe<sup>1,2</sup>, Kenichi Inoue<sup>1</sup>, Yasuhide Furuta<sup>1,2</sup>

<sup>1</sup>Animal Resource Development Unit, <sup>2</sup>Genetic Engineering Team, RIKEN CLST, Kobe, Japan

**P-92 Streamlining the generation of knockout and knockin mice using the CRISPR-Cas system**

Suman Komketi, Michela Ascolani, Alessandra Pisaniello, Antonella Vecchio,

Marco Riposati, T. Neil Dear

EMBL Rome, Rome, Italy

**P-93 Improved Oviduct Transfer Surgery for Genetically Modified Rat Production**

Laura J. Lambert<sup>1</sup>, Larry W. Johnson<sup>1</sup>, Daniel Kennedy<sup>1</sup>, Joan Cadillac<sup>2</sup>,

Robert A. Kesterson<sup>1</sup>,

<sup>1</sup>Department of Genetics, the University of Alabama at Birmingham, Birmingham, USA; <sup>2</sup>Animal Resources Program, the University of Alabama at Birmingham, Birmingham, USA

### **P-94 Employing single-stranded DNA oligonucleotides for the high throughput production of conditional knockout alleles in mice**

Angelina Gaspero<sup>1</sup>, Ying Wang<sup>1</sup>, Isabel Lorenzo<sup>1</sup>, Lan Liao<sup>1</sup>, John Seavitt<sup>1</sup>, Mary Dickenson<sup>1</sup>, Arthur Beaudet<sup>1</sup>, Francisco DeMayo<sup>2</sup>, Jianming Xu<sup>1</sup>, Jason D Heaney<sup>1</sup>, Denise Lanza<sup>1</sup>,  
<sup>1</sup>Baylor College of Medicine, Houston, TX, United States; <sup>2</sup>National Institute of Environmental Health Sciences, Research Triangle Park, NC, United States

### **P-95 Development of KCNQ1 knockout golden Syrian hamsters as a model of Jervell and Lange-Nielsen syndrome**

Rong Li<sup>1</sup>, Justin Lichter<sup>2</sup>, Jianjun Wang<sup>3</sup>, Jinxin Miao<sup>1</sup>, Derek Dossdall<sup>2</sup>, Ravi Ranjan<sup>2</sup>, Xi Lin<sup>3</sup>, Don Sinex<sup>4</sup>, Zhongde Wang<sup>1</sup>  
<sup>1</sup>Dept. Animal, Dairy, and Veterinary Sciences, Utah State University, Logan, UT; <sup>2</sup>Division of Cardiovascular Medicine, University of Utah, Salt Lake City, UT; <sup>3</sup>Dept. Otolaryngology-Head & Neck Surgery, Emory University School of Medicine, Atlanta, GA; <sup>4</sup>COMDDE, Utah State University, UT

### **P-96 A sensitive method for detecting multiple mutations generated by CRISPR/Cas9 genome editing**

Don Liu, Gregory Martin, Uma Saha, Cheryl DiCarlo  
Charles River Laboratories, Wilmington, MA, United States

### **P-97 Development of a mouse model suitable for in vivo genome editing efficiency studies**

Hiroshi Miura<sup>1,2</sup>, Yutaka Inagaki<sup>2</sup>, Channabasavaiah B Gurumurthy<sup>3</sup>, Masahiro Sato<sup>5</sup>, Masato Ohtsuka<sup>1</sup>  
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### **P-98 Cas9 RNA-guided nuclease – efficient model generation by microinjection and electroporation**

Lauryl M. J. Nutter<sup>1,2</sup>, Marina Gertsenstein<sup>1</sup>, Janet Rossant<sup>3</sup>  
<sup>1</sup>Model Production Core, The Centre for Phenogenomics, Toronto, Canada; <sup>2</sup>Translational Medicine, The Hospital for Sick Children, Toronto, Canada; <sup>3</sup>Developmental and Stem Cell Biology, Peter Gilgan Centre for Research and Learning, The Hospital for Sick Children, Toronto, Canada; Department of Molecular Genetics, University of Toronto, Canada

### **P-99 Opportunity Knocks**

Nicola Osborne  
Responsible Research in Practice, West Sussex, Horsham, UK



TT2017

## 14TH TRANSGENIC TECHNOLOGY MEETING

OCTOBER 1-4 | SNOWBIRD RESORT, SALT LAKE CITY, UTAH, USA

### **P-100 Effects of voltage strength on development and quality of electroporated porcine embryos**

Katsutoshi Nishio, Fuminori Tanihara, Thanh-Van Nguyen, Toshiki Kuniyama, Takeshige Otoi  
Bioscience and Bioindustry, Tokushima University, Myozai-gun, Tokushima, Japan

### **P-101 Analysis of mouse blastocysts to assess in vivo activity of sgRNAs**

Elena McBeath<sup>1</sup>, Chad Smith<sup>2</sup>, Jun-ichi Abe<sup>1</sup>, Keigi Fujiwara<sup>1</sup>, Jan Parker-Thornburg<sup>2</sup>  
<sup>1</sup>Department of Cardiology, MD Anderson Cancer Center, Houston TX, USA; <sup>2</sup>Department of Genetics, MD Anderson Cancer Center, Houston TX, USA

### **P-102 Management of mouse sperm freezing and quality control**

Nicole Kuepper, Julia Mock, Sonja Kropp, Klaus Pfeffer  
Heinrich-Heine-University Duesseldorf, Duesseldorf, Germany

### **P-103 Generating Murine Models in NZBWF1/J Background for Systemic Lupus Erythematosus**

Tuija M Alcantar, Jinjie Li, Natasha O'Neil, Mandy Thayer, Lisa Lima, Linda Ta, Xin Y Rairdan,  
Genentech, Inc., South San Francisco, CA, United States

### **P-104 CRISPR and the 3Rs - reducing animal numbers in high-throughput mouse production**

Edward Ryder, Brendan Doe, Joanna Bottomley, Katharina Boroviak, Graham Duddy, Diane Gleeson, Hannah Wardle-Jones, Ramiro Ramirez-Solis, James Bussell, Sanger Mouse Genetics Project  
Wellcome Trust Sanger Institute, Hinxton, Cambridge, United Kingdom

### **P-105 Non-Surgical Embryo Transfer in Rats; A 3Rs Refinement for Assisted Reproductive Technology**

Barbara J Stone, Kendra H Steele, Sarah J Srodulski  
ParaTechs Corporation, Lexington, KY, United States

### **P-106 Optimization of CRISPR reagent delivery using the zygote electroporation of nucleases (ZEN) method**

Wenhua Li, Timothy Dahlem, Susan Tamowski  
University of Utah, Salt Lake City, Utah, United States

### **P-107 Assessing the Effects of Filtering CRISPR Reagents through a Centrifuge Tube Filter Prior to Microinjection**

Michael Woods, Brendan Doe, Caroline Sinclair, Ellen Brown, Katharina Boroviak, Graham Duddy, Ed Ryder  
Wellcome Trust Sanger Institute, Hinxton, Cambridge, United Kingdom

### **P-108 CRISPR/Cas9 Gene Editing in Mice**

Zhenjuan Wang, Ying Chen, Laurie R Chen, Sarah J Johnson, Lin Wu  
Harvard University, Cambridge, MA, United States

### **P-109 Improve Knock in Efficiency through CRISPR/CAS9 Guide RNA Rescue**

Dongshan Yang, Jun Song, Jie Xu, Jinxue Ruan, Jifeng Zhang, Yuqing Eugene Chen  
University of Michigan Medical Center, Ann Arbor, Michigan, United States

### **P-110 Recombineering Bacterial Artificial Chromosome Transgenes**

Michael Zeidler, Thomas L Saunders  
University of Michigan, Ann Arbor, MI, United States

### **P-111 Towards an optimized workflow for CRISPR/Cas mediated mouse transgenesis**

Simon Tröder, Sonja Assenmacher, Kerstin Weisheit, Patrick Jankowski, Branko Zevnik  
CECAD, University of Cologne, Cologne, Germany

### **P-112 Production of immunodeficient rabbits by multiplex embryo transfer and multiplex gene targeting**

Jun Song<sup>1</sup>, Dongshan Yang<sup>1</sup>, Jinxue Ruan, Jifeng Zhang, Yuqing Eugene Chen, Jie Xu  
University of Michigan Medical Center, Ann Arbor, Michigan, United States; <sup>1</sup>These authors contribute equally to the work

### **P-114 Refining Chimera Production via Microinjection**

Sophie Wood, Daniel Snell  
Francis Crick Institute, London, UK

### **P-115 Development of Cre driver rat strains for creating conditional and physiologically relevant models of human diseases**

James Jinping Luo, Jinling Li, Ruby Yanru Chen-Tsai  
Applied StemCell, Inc. Milpitas, CA, USA

### **P-116 How To Generate Mouse Models: Right Tool For The Right Job**

Roger Caothien, Charles Yu, Lucinda Tam, Sobha Thamminana, Anu Sebin, Ana Carolina Magalhaes Mota, Soren Warming, Merone Roose-Girma  
Genentech, San Francisco, CA, USA